ORIGINAL RESEARCH

Inulin Can Alleviate Metabolism Disorders in ob/ob Mice by Partially Restoring Leptin-related Pathways Mediated by Gut Microbiota

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Received 25 October 2018; revised 13 March 2019; accepted 19 March 2019
Available online 23 April 2019
Handled by Kang Ning

Abstract  Inulin has been used as a prebiotic to alleviate glucose and lipid metabolism disorders in mice and humans by modulating the gut microbiota. However, the mechanism underlying the alleviation of metabolic disorders by inulin through interactions between the gut microbiota and host

Peer review under responsibility of Beijing Institute of Genomics, Chinese Academy of Sciences and Genetics Society of China.

https://doi.org/10.1016/j.gpb.2019.03.001
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cells is unclear. We use ob/ob mice as a model to study the effect of inulin on the cecal microbiota by 16S rRNA gene amplicon sequencing and its interaction with host cells by transcriptomics. The inulin-supplemented diet improved glucose and lipid metabolism disorder parameters in ob/ob mice, alleviating fat accumulation and glucose intolerance. The α diversity of gut microbial community of ob/ob mice was reduced after inulin treatment, while the β diversity tended to return to the level of wild type mice. Interestingly, Prevotellaceae UCG 001 (family Prevotellaceae) was obviously enriched after inulin treatment. A comparative analysis of the gene expression profile showed that the cecal transcriptome was changed in leptin gene deficiency mice, whereas the inulin-supplemented diet partially reversed the changes in leptin gene-related signaling pathways, especially AMPK signaling pathway, where the levels of gene expression became comparable to those in wild type mice. Further analysis indicated that Prevotellaceae UCG 001 was positively correlated with the AMPK signaling pathway, which was negatively correlated with markers of glycolipid metabolism disorders. Our results suggest that the inulin-supplemented diet alleviates glucose and lipid metabolism disorders by partially restoring leptin related pathways mediated by gut microbiota.

Introduction

Inulin is a common prebiotic defined as a non-digestible dietary fiber that supports the growth of probiotics [1]. To be more specific, it can stimulate the growth of bifidobacteria in the intestine [2]. Therefore, inulin has been used to regulate gut microbiota-related disorders, such as metabolic disorders, allergies, and inflammatory bowel disease (IBD) [3–6]. To date, only one prebiotic has been identified by European Food Safety Authority: inulin improves bowel function [7]. In addition, the probiotic effect of inulin, such as changing the gut microbiota of obese individuals, increasing the abundances of bifidobacteria and Akkermansia muciniphila in obese individuals and improving metabolic disorders, have been widely reported [5,8–11]. Inulin can be enzymatically hydrolyzed by some bacteria to produce short-chain fatty acids (SCFAs) in the colon, which can bind to G protein-coupled receptors (e.g., GPR41/43) expressed on the intestinal epithelial cell membrane and then stimulate Glucagon-like peptide-1 (GLP-1) production. As a consequence, GLP-1 has a chronic effect on the host energy metabolism system [12,13].

Several metabolites produced by the gut microbiota (e.g., SCFAs and lipopolysaccharide (LPS)) can affect host gene expression profile and regulate host energy balance involving free fatty acid receptor (FFA) 2/3, fasting-induced adipocyte factor (Fiaf), and adenosine monophosphate (AMP)-activated protein kinase (AMPK) [13–16]. Additionally, SCFAs can regulate the secretion of gut peptides (e.g., GLP-1 and Peptides YY (PYY)) by recognizing receptors on the surface of intestinal cells. These gut peptides have positive effects on reducing food intake and improving glucose metabolism [17]. Conversely, LPS, as a type of pathogen-associated molecular patterns (PAMPs), can trigger low-grade inflammation, which leads to the occurrence of metabolism disorders [18].

Obesity is a typical metabolic disorder, and the gut microbiota is regarded as an important body part that may provide additional contributions to obesity besides the contributions of the host genotype and lifestyle. Previous studies have provided solid evidence that the gut microbiota can affect host energy intake [15,19].

AMPK as a key protein kinases of host energy status, plays an essential role in host energy balance. It can regulate the expression of adipokines, which are involved in body weight control, appetite control, and maintenance of metabolism homeostasis [20]. In general, the role of AMPK in liver, fat, and skeletal muscle tissue is critical, and the activity of AMPK can be tuned by different factors such as drug intervention.

We hypothesize that the AMPK signaling pathway can also play an important role in modulating gene expression of cecal tissue after the inulin-supplemented diet intervention and has effects on other signaling pathways related to obesity. To understand the mechanism underlying the improvement in glycolipid metabolism disorders by inulin in obese individuals, ob/ob mice were selected to seek the effect of the inulin-supplemented diet on the cecal microbiota and its possible influence on the host cecal gene expression profile.

Results

The inulin-supplemented diet can improve metabolic disorder-related symptoms in genetically obese mice

To investigate the effect of inulin treatment on glycolipid metabolism disorders in genetically obese mice, we examined the body weight, daily food intake, liver weight and glycolipid metabolism related parameters. The initial body weight (Figure S1), daily food intake, liver index, serum total cholesterol (TC), TC/high-density lipoprotein cholesterol (HDL-C) ratio and the area under the curve (AUC) of the Intraperitoneal glucose tolerance test (IPGTT) of the ob/ob mice (ob/ob) were significantly elevated compared with those of wild type mice (P < 0.05, one-way analysis of variance (ANOVA) followed by the Tukey post hoc test, Figure 1A–F), indicating severe glucose intolerance and dyslipidemia. In contrast, ob/ob mice fed the inulin-supplemented diet (ob/ob inulin) for 4 weeks showed improved metabolic parameters. After inulin intervention, the treated group showed no statistically significant difference in body weight change compared with the ob/ob group (Figure S2), but had a significantly lower daily food intake (P < 0.05, one-way ANOVA followed by the Tukey post hoc test, Figure 1A), serum TC, and TC/HDL-C ratio (P < 0.05, one-way ANOVA followed by the Tukey post hoc test, Figure 1C, D). Additionally, the AUC of IPGTT was lowered, indicating a significant improvement in glucose tolerance (P < 0.05, one-way ANOVA followed by the Tukey post hoc test, Figure 1E, F). Subsequently, we measured the Glp-1 mRNA level in colonic tissue. The inulin-supplemented diet could rise the Glp-1 mRNA level in ob/ob inulin, meaning that inulin restored glucose homeostasis in ob/ob mice (P < 0.05,
one-way ANOVA followed by the Tukey post hoc test, Figure 1G), which was not reported previously. According to liver tissue staining (Figure 1H), ob/ob mice exhibited severe hepatic steatosis, which was alleviated after inulin treatment. Overall, the above results indicate that inulin has a beneficial effect that ameliorates glycolipid metabolism disorders in ob/ob mice.

The ob/ob mice fed the inulin-supplemented diet show apparent changes in gut microbial structure.

To explore the effect of inulin treatment on the gut microbiota of ob/ob mice, we performed 16S rRNA sequencing of the cecal contents. We used the 16S rRNA gene amplicon sequencing method (V3-V4 region) and generated 3,179,284 reads for a total of 18 samples, with an average of 176,627 ± 22,171 reads per sample. Compared with the genetically obese mice, the treated group had slightly fewer operational taxonomic units (OTUs) ($P = 0.063$, one-way ANOVA followed by the Tukey post hoc test, Figure 2A) and a significantly lower $\alpha$ diversity index (Shannon and Simpson indexes) ($P < 0.05$, one-way ANOVA followed by the Tukey post hoc test, Figure 2B, C).

Principal coordinates analysis (PCoA) of the weighted UniFrac distances analysis for the gut microbiota of the three mouse groups showed that the wild type and ob/ob groups were clearly clustered into two separate groups, while the values for the ob/ob inulin mice were clustered between the two groups.
groups. However, the three groups shared some overlapping regions (Figure S3). In summary, we did not detect significant differences in bacterial diversity between wild type and ob/ob mice, while there was a slightly lower bacterial diversity in ob/ob inulin mice.

The phylum Bacteroidetes was dominant among the 9 phyla present in the gut microbiota from the three groups of mice, and the ratio of Firmicutes/Bacteroidetes was increased in ob/ob mice over wild type group, but lower in the ob/ob inulin group compared with ob/ob mice (Figure S4). The gut

Figure 2  Inulin modifies the composition of the cecal microbiota in ob/ob mice
A. OTU number. B. Shannon index of microbiota. C. Simpson index of microbiota. D. LDA scores of differentially abundant taxa between the wild type and ob/ob mice using the LEfSe method. E. LDA scores of differentially abundant taxa between the ob/ob and ob/ob inulin mice using the LEfSe method. F. Relative abundance of Prevotellaceae UCG 001. G. Relative abundance of Alistipes, Anaerotruncus, Intestimonas, and Family XIII UCG 001. OTU, operational taxonomic unit. Data were analyzed using one-way ANOVA followed by the Tukey post hoc test for A–C, and Kruskal–Wallis sum-rank test and Wilcoxon rank-sum test for D, E. n = 6 per group. *p < 0.05.
microbiota in obese individuals has usually shown an increased Firmicutes/Bacteroidetes ratio [21]. Therefore, the decreased Firmicutes/Bacteroidetes ratio of ob/ob inulin means that this feature in obesity could be reversed by the inulin-supplemented diet.

Next, to identify the changes in specific bacterial taxa after the inulin-supplemented diet intervention, we utilized the linear discriminant analysis (LDA) effect size (LEfSe) to compare the cecal microbiota composition between the ob/ob and ob/ob inulin groups. At the genus level, LDA score was selected to discriminate specific taxa in different groups. Compared with the wild type group, the ob/ob mice had a higher abundance of Alistipes, Anaerotruncus, Roseburia, Candidatus Saccharimonas, Lachnospiraceae UCG 001, Intestinimonas, Turicibacter, and Family XIII UCG 001 but a lower abundance of Prevotellaceae UCG 001, Parasutterella, Parabacteroides, Erysipelotrichaceae, and Bilophila (Figure 2D). Correspondingly, Alistipes, Anaerotruncus, Desulfovibrio, Lactobacillus, Butyricicoccus, Intestinimonas, Ruminococcaceae UCG 001, and Ruminococcaceae UCG 005 were enriched in the ob/ob group compared to wild type (762 upregulated genes, including 242 genes with more than doubled expression levels; 446 downregulated genes, including 138 with more than halved expression levels, P < 0.05, Wald test, Figure S6). To further investigate the biological signaling pathways involved in the above DEGs, we used the Kyoto Encyclopedia of Genes and Genomes (KEGG) to do the enrichment analysis. We found that there were 232 signaling pathways involved in the above DEGs between the ob/ob and wild type mice.
and wild type groups. Subsequently, according to the results of the correlation analysis, we obtained 50 candidate signaling pathways, which had strong correlations with the 5 metabolic parameters. These signaling pathways were mainly involved in certain KEGG categories, such as Environmental information processing, Organismal systems, and Metabolism. This result indicated that leptin gene deficiency in ob/ob mice affected some metabolism-related signaling pathways.

The inulin-supplemented diet restores certain metabolism-related signaling pathways in ob/ob mice

We then investigated the effect of inulin treatment on the cecal transcriptome of obese mice. As the above results indicated that some metabolism-related signaling pathways of the cecum were changed in leptin gene deficiency mice, we continued to analyze the cecal transcriptome data after the inulin-supplemented diet. In total, there were 362 DEGs between ob/ob inulin and ob/ob (216 upregulated genes, including 42 genes with more than doubled expression; 146 downregulated genes, including 34 genes with more than halved expression levels, \( P < 0.05 \), Wald test, Figure S7). We found that 140 signaling pathways were involved in these DEGs. Using the same correlation analysis method as above, we selected 62 candidate signaling pathways. Finally, comparisons of the candidate pathways between ob/ob vs. wild type and ob/ob inulin vs. ob/ob identified 8 signaling pathways for which the enrichment score was changed due to leptin gene deficiency but restored by the inulin-supplemented diet (Figure 3A). In detail, the AMPK signaling pathway, Sphingolipid signaling pathway, Dopaminergic synapse, and Glycine/serine/threonine metabolism were downregulated, and in contrast, Pyruvate metabolism, Glycolysis/Gluconeogenesis, Arachidonic acid metabolism, and Thyroid hormone synthesis were elevated in the ob/ob group compared with the wild type group (\( p < 0.05 \), one-way ANOVA followed by the Tukey post hoc test, Figure 3B). Furthermore, the change in the enrichment score of these 8 signaling pathways was reversed after the inulin-supplemented diet (Table S1).

Correlations among the bacteria, signaling pathways, and metabolic parameters

Here, to further analyze the association between the cecal microbiota and the cecal transcriptome, we performed correlation analyses comparing the 5 bacterial genera and 8 signaling pathways with 5 metabolic parameters. These correlation analyses showed that Prevotellaceae UCG 001 had a significant negative correlation with liver index and TC/HDL-C. However, Alistipes, Anaerotruncus, and Family XIII UCG 001 were positively correlated with these parameters. Additionally, we found that the AMPK signaling pathway, Sphingolipid signaling pathway, Dopaminergic synapse, and Glycine/serine/threonine metabolism were negatively correlated with the 5 metabolic parameters, while Pyruvate metabolism, Glycolysis/Gluconeogenesis, Thyroid hormone synthesis, and Arachidonic acid metabolism had positive correlations with the metabolic parameters (Figure 4A).

Obviously, the gut microbiota could be affected by both leptin gene deficiency and inulin-supplemented diet. We performed correlation analyses between the 5 bacterial genera and 8 signaling pathways. The results showed that there was a significant positive correlation between the enrichment score of the AMPK signaling pathway and Prevotellaceae UCG 001, and the AMPK signaling pathway was negatively associated with the other 4 bacterial genera. However, the abundance of Prevotellaceae UCG 001 had significant positive correlations with the enrichment scores of the AMPK signaling pathway, Sphingolipid signaling pathway, and Dopaminergic synapse. Conversely, Alistipes had significant positive correlations with Pyruvate metabolism, Glycolysis/Gluconeogenesis, Thyroid hormone synthesis, and Arachidonic acid metabolism, which were negatively correlated with the abundance of Prevotellaceae UCG 001 (Figure 4B).

Discussion

Disorder of the gut microbiota has been considered one of the reasons for metabolic disorders. Gut microbes can regulate gut

**Figure 4 Correlations among gut bacteria, signaling pathways, and metabolic parameters**

A. Heatmap of the Spearman correlations between 5 genera, 8 signaling pathways and 5 metabolic parameters. B. Heatmap of the Spearman correlations between 5 genera and 8 signaling pathways. TC, total cholesterol; HDL-C, high-density lipoprotein cholesterol; AUC, area under the curve; IPGTT, intraperitoneal glucose tolerance test. \( n = 6 \) per group. *\( P < 0.05 \).
micro-ecology through cell surface molecules and/or their metabolites, thereby affecting the host metabolic system, immune system, and nervous system [19,22–24]. Although prebiotics can exert positive effects on the maintenance of host metabolic homeostasis, which are mainly mediated by gut microbiota [7], there are few reports on the effects of prebiotics on the intestinal gene expression [5,11]. Genetically obese mice and lean mice have marked differences in several metabolic parameters and the gut microbiota [4]. Here, we reveal that the reason for the prebiotic inulin alleviating obesity-related glucose and lipid metabolism disorders in leptin gene deficiency mice may be closely related to the restoring of certain metabolism-related pathways.

In this study, after 4 weeks of the inulin-supplemented diet in ob/ob mice, significant alleviation of metabolic disorders was observed. Compared to the wild type group, the ob/ob group had a 71% higher body weight at 10 weeks of age, and their daily food intake increased by approximately 29%, which indicates that, due to the lack of functional leptin, the genetically obese mice lost their appetite control, but this status was improved after inulin treatment. We observed increases in serum TC, TC/HDL-C, and the AUC of IPGTT, which indicated that the glucose and lipid metabolism disorders were severe, as evidenced by the liver histopathology and liver index. The decreases in the above metabolic parameters after the inulin-supplemented diet suggest that the metabolic disorders in ob/ob mice were alleviated. Moreover, non-digestible fermentable dietary fibers can exert antidiabetic effects by increasing the secretion of GLP-1, which is secreted by intestinal epithelial cells, and this hormone can take part in maintaining blood glucose homeostasis [25]. We therefore measured the Glp-1 mRNA level in the colon and found that this gene was significantly elevated after inulin treatment, suggesting that the maintenance of blood glucose homeostasis by inulin-supplemented diet was probably through the increased expression of Glp-1 gene.

The abundance of the family Prevotellaceae (phylum Bacteroidetes) was increased by inulin, and Prevotellaceae UCG 001 belonging to this family was enriched in the ob/ob inulin group. Prevotella possesses enzymes that can degrade cellulose and xylan [26]. As succinate-producing bacteria, Prevotella can participate in the degradation of inulin [27]. Dietary fiber facilitates the colonization of Prevotella in the gut, which in turn improves glucose metabolism [28]. A study utilizing wheat-type cereal products showed that cereals could alter the gut microbiota composition of infants, especially by increasing the family Prevotellaceae, which can degrade cellulose [29]. In a mouse model study, whole wheat oats altered the gut microbiota, increased the abundance of Prevotella and then enhanced insulin function and plasma lipid regulation [30]. In agreement with these observations, the abundance of Prevotellaceae UCG 001 was negatively correlated with the blood glucose- and lipid-related parameters, especially TC/HDL-C and liver index. Therefore, the elevation in abundance of Prevotellaceae UCG 001 might be closely related to the beneficial effect of the inulin-supplemented diet. Thus, we speculated that bacteria of this genus might affect the host glucose and lipid metabolism through the production of secondary metabolites, such as SCFAs.

In contrast with the lean mice, the abundance of Alistipes in the gut of the ob/ob mice was significantly increased but was reduced after the inulin-supplemented diet. Long-term ingestion of high-sugar foods can lead to enrichment of this genus in the human intestinal tract, as well as in animal experiments [31,32]. The abundance of this genus, which contains pro-inflammatory bacteria, was significantly increased in the gut of obese individuals in Japan [33]. In this study, Alistipes was significantly positively correlated with serum TC, TC/HDL-C levels, and the AUC of IPGTT, which were identified as indicators of glucose and lipid metabolism disorders. Therefore, we speculated that when Alistipes was inhibited after the inulin-supplemented diet, its promotion of metabolic disorders decreased.

Arachidonic acid, an unsaturated fatty acid, is positively related to pro-inflammatory cytokines, but inversely correlated with Akkermansia and exacerbates non-alcoholic steatohepatitis (NASH) by promoting the growth of pro-inflammatory bacteria, enhancing the inflammatory response, reducing the abundance of butyrate-producing bacteria, and inducing insulin resistance [34,35]. A high-fat diet can induce thyroid dysfunction in rodent models, and that the thyroid volume is significantly increased in obese individuals, in contrast to healthy ones [36,37]. Here, the enrichment scores for Arachidonic acid metabolism and Thyroid hormone synthesis were restored to relatively lower levels after the inulin-supplemented diet.

Importantly, a significant increase in the enrichment score for the AMPK signaling pathway was exhibited after inulin supplementation. In vivo, the activation of AMPK signaling can stimulate glucose uptake, phosphorylation of acetyl coenzyme A carboxylase, hepatic glycogenolysis, lactate production and fatty acid oxidation, and correspondingly, it inhibits hepatic gluconeogenesis, cholesterol and fatty acid synthesis [38,39]. AMPK has been well demonstrated that it can act as a crucial sensor in regulation of glycolipid metabolism. For example, AMPK influences energy intake, utilization, and storage in the skeletal muscle, heart, adipose tissue, liver, pancreatic beta cells, and brain by regulating dietary intake and substrate metabolism [20,40]. The lactate and butyrate produced by certain intestinal bacteria can increase AMPK activity, which in turn increases fatty acid oxidation and energy expenditure [41]. Metformin and thiazolidinediones, widely used drugs for clinical treatment for diabetes, can promote fatty acid oxidation, and inhibit the activity of lipogenesis-related enzymes by activating AMPK in liver cells [42,43]. Germ-free mice can resist the obesity induced by high-fat diets mainly by increasing AMPK phosphorylation in the liver and skeletal muscle. Phosphorylated AMPK can promote fatty acid oxidation in peripheral tissues [44]. Fat deposits in the liver and skeletal muscle are also regulated by intestinal bacteria through AMPK [45]. Obviously, AMPK plays a vital role in maintaining glycolipid homeostasis, but previous studies involving AMPK mainly focused on the alteration of this protein in the skeletal muscle and liver. In our study, in contrast to lean mice, the enrichment score of the AMPK signaling pathway in the cecum of ob/ob mice was reduced by 36%, and the ob/ob inulin mice had a 66% higher score than the ob/ob mice. The enrichment score of the AMPK signaling pathway was negatively correlated with liver index, daily food intake, serum TC, TC/HDL-C, and the AUC of IPGTT, but Glycolysis/Gluconeogenesis and Thyroid hormone synthesis showed positive correlations with these parameters. Furthermore, the results showed a high correlation between Prevotellaceae UCG 001 and the enrichment score of the AMPK signaling pathway.
In contrast, the abundance of *Alistipes* was significantly negatively correlated with this pathway. Interestingly, to date, correlations between the AMPK signaling pathway and these different bacteria have not been reported. Additionally, our findings emphasized the close interrelation between the cecal microbiota and the cecal transcriptome.

AMPK could be used as a potential target for the treatment of diabetes and obesity. AMPK activation in the liver can affect whole-body metabolism and lead to improvements in circulating levels of glucose and lipid [38,46], and we found that this effect was not only present in the liver and skeletal muscle, but also in the cecum.

Compared with *ob/ob* group, the relative expression of colonic Glp-1 mRNA was significantly increased in the *ob/ob* inulin group. As one of hormones produced by the gut, GLP-1 is a well-studied anorexigenic hormone. GLP-1 plays an important role in glucose homeostasis, gastrointestinal motility and appetite suppression [47–49]. SCFAs can stimulate the expression of GLP-1 from enteroendocrine L-cells, and the release of this hormone reduces food intake and maintains glucose homeostasis [50].

Overall, leptin gene deficiency led to metabolic disorders and inhibition of the AMPK signaling pathway in genetically obese mice. We speculated that the composition of cecal microbiota could be affected by inulin-supplemented diet. In particular, *Prevotellaceae UCG 001*, which produces SCFAs by degrading inulin, was increased. Then, SCFAs recognized and bound to GPR41/43 on the cell membrane of the cecal epithelium, triggering intracellular AMPK phosphorylation and the subsequent changes in downstream metabolic functions (such as promotion of fatty acid oxidation, glycolysis, and inhibition of hepatic fatty acid synthesis, cholesterol synthesis, and gluconeogenesis). Eventually, the host metabolic disorders were ameliorated (Figure 5).

**Conclusions**

Prebiotic effect of inulin might be involving in the host transcriptome changing mediated by altering the gut microbiota. Based on the results of 16S rRNA gene amplicon analysis and transcriptomic analysis, we explored the complex interplay...
between the cecal microbiota and host gene expression profile. We first reported the effect of prebiotics on the cecal transcriptome in genetically obese mice. Giving inulin (10 g/kg/day) to ob/ob mice can alleviate glucose and lipid metabolism disorders, increase the abundance of Prevotellaceae UCG 001, and reduce the abundance of Allistipes. Moreover, we found that the AMPK signaling pathway in cecal tissue was upregulated after inulin supplementation. However, further studies are required to reveal the precise mechanism(s) behind these effects. Our results suggest that the AMPK signaling pathway not only plays a pivotal role in regulating metabolism in adipose tissue, liver, and skeletal muscle but also plays a vital role in the metabolic regulation mediated by gut microbiota. Furthermore, similar to the key role of the AMPK signaling pathway in the liver and skeletal muscle, the alteration of this pathway in the cecum may also play an indispensable role in the interaction between the cecal microbiota and the host metabolism system. Overall, signaling pathways related to energy metabolism in the gut are not negligible.

Materials and methods

Animals

Six-week-old male C57BL/6J mice were used in the experiment. C57BL/6J and ob/ob mice (C57BL/6J background; Huafukang Bio-Technique Co., Ltd, Beijing, China) were placed in a controlled environment (12 h sunlight cycle; lights off at 6 p.m.) with two mice in each cage given ad libitum access to feed and water. Upon delivery, the mice were acclimated for a period of 2 weeks, during which they were fed a normal chow diet (13.5% calories from fat; Vital River Laboratory Animal Technology Co., Ltd, Beijing, China). C57BL/6J mice were fed a normal chow diet (n = 6), while ob/ob mice were fed a normal chow diet supplemented (n = 6) or not (n = 6) with inulin (D9082A0044, Purity > 90%, BBI Life Sciences Co., Ltd, Shanghai, China) at 10 g/kg/day through drinking water. The duration of the intervention was 4 weeks. Body weight was measured once weekly, and food and water intake were recorded daily.

ob/ob mice, which are deficient in the leptin gene (Lepob/Lepob), are one of the most common models for diabetes and obesity studies. There is a C→T mutation in C57BL/6J ob/ob mice that results in a change of an arginine at position 105 to a stop codon, which results in the synthesis of a truncated protein that is degraded in the adipocyte [51]. This gene deficiency can induce variations in key parameters such as glucose intolerance [52].

All animal experiments were approved by the Ethics Committee of the Institute of Microbiology, Chinese Academy of Sciences (IMCAS) and followed the Declaration of Helsinki (SQIMCAS2017009).

IPGTT

For the glucose tolerance test, an IPGTT was carried out at the end (week 10) of the treatment. The mice were fasted for 12 h and then injected subcutaneously with glucose (2.0 g/kg body weight). The measurement of blood glucose with a blood glucose meter (Accu-Check, Roche, Switzerland) was conducted with tail bleeding at −30, 0, 15, 30, 60, 90, and 120 min before or after the intraperitoneal glucose load.

Tissue sampling

At the end of the treatment period, the mice were anesthetized by an intraperitoneal injection of pellitobarbitalum matricum (80 mg/kg body weight). Serum samples were collected for further detection. Cervical dislocation was used to killed mice. Epididymal fat tissue, the liver, the spleen and intestinal segments (cecum and colon) were precisely dissected and weighed. Then, these tissues and serum samples were immediately submerged in liquid nitrogen and transferred to −80 °C for preservation.

Histological analyses

Freshly isolated livers from all three groups were rapidly immersed in 4% formaldehyde for 24 h at ambient temperature. The tissues were then fixed in absolute ethyl alcohol for 24 h, and embedded in paraffin wax and cut into 5-μm-thick tissue slices. The slices were stained with hematoxylin/eosin (H&E) and detected by a DM2000 light microscope (Leica Microsystems GmbH, Wetzlar, Germany) at 100× magnification.

Biochemical analyses

The whole blood was coagulated at room temperature for 30 min, and centrifuged at 4 °C and 3000 x g for 10 min to collect serum. The serum was stored at −80 °C. The levels of serum TC, triglycerides (TG), HDL-C and low-density lipoprotein cholesterol (LDL-C) were measured using a commercial detection kit (NIJCBIO Co., Ltd, Nanjing, China) according to the kit instructions. Serum interleukin-1α (IL-1α), adiponectin (ADPN), and insulin were quantified using commercial enzyme-linked immunosorbent assay (ELISA) kits (NIJCBIO Co., Ltd, Nanjing, China) according to the kit instructions. Automated microplate reader was used to detect the absorbance values at 450 nm within 30 min.

Microbiota analysis

After mice were dissected, the cecal contents were collected and stored at −80 °C. Microbial DNA was extracted from the cecal content using a TIANamp Stool DNA kit (TIANGEN Bio-Tech Co., Ltd, Beijing, China) according to the manufacturer’s protocol, including a bead-beating step. The V3-V4 region of the bacterial 16S ribosomal RNA gene was amplified by PCR (95 °C for 3 min, followed by 25 cycles at 95 °C for 30 s, 55 °C for 30 s, 72 °C for 30 s, and a final extension at 72 °C for 5 min) using primers F3 (CCTACGGGNBGG-CTACCT) and R4 (GGTTACCCAGCMGCCGCGGTAA). High-throughput sequencing was performed utilizing the Illumina HiSeq 2500 PE250 platform to detect the 16S rRNA amplicons according to standard protocols. Paired-end reads from sequencing were merged utilizing Fast Length Adjustment of SHort reads (FLASH) [53]. The fastq_quality_filter (-p 90 -q 25 -q33) belonging to FASTX Toolkit 0.0.14 was used to filter low quality reads. USEARCH 64 bit v8.0.1517 was utilized to
remove chimaeric reads. Normalizing the reads counts of each sample was based on the minimum size of samples by random subtraction. The UCLUST algorithm was used to align OTUs at 97% identity. The SILVA 16S rRNA database v128 was utilized to taxonomically classify OTUs. The Quantitative Insights Into Microbial Ecology (QIIME) was applied to calculate the α and β diversities, which were computed based on weighted and unweighted UniFrac distance matrices [54]. The significantly differential species between groups were generated by the LEfSe method [55].

RNA-seq analysis

TRIZol reagent (Invitrogen, Thermo Scientific, MA, USA) was used to isolate total RNA of cecal samples following the instructions. Each sample required 4 μg of total RNA to construct a cDNA library. The library was constructed using the KAPA Stranded mRNA-Seq Kit for the Illumina platform (KAPA-BIO, Boston, USA) according to the manufacturer’s protocols. The library preparations were sequenced on an Illumina HiSeq PE150 platform.

The low quality reads and adaptor sequences were trimmed with Trimmomatic [56]. Clean reads were aligned to mm10 reference genome with Hisat2 [57]. Gene expression levels were calculated by counting the overlap of reads on each gene with HT-seq [58] and normalized as reads per kilobase per million mapped reads (RPKM) with the gene annotation file from Ensembl (release 87) and the DESeq2 package in R [59]. In addition, the DESeq2 package was applied to calculate the differentially expressed genes (DEGs). Functional enrichment in Gene Ontology (GO) and KEGG was performed with the GOstats package [60]. The enrichment scores of pathways, which were used to compare the differences in pathways between two groups, were calculated by the RPKM values of the DEGs contained within each pathway.

Availability of data

The datasets supporting the conclusions of this article are available in the NCBI repository (https://www.ncbi.nlm.nih.gov/); SRA accession SRP154971 for 16S rRNA gene amplification sequencing data; SRA accession SRP155684 for RNA sequencing data.

Real-time qPCR analysis

The FastQuant RT kit (TIANGEN BIO, Beijing, China) was applied to prepare cDNA according to the kit instructions. Real-time qPCR assays in triplicate were performed using the Applied Biosystems® 7500 Real-Time PCR System (Thermo Scientific, MA, USA) with KAPA SYBR FAST qPCR kit Master Mix (KAPA-BIO, Boston, USA). The expression of target genes in mice was normalized to that of the gene encoding glyceraldehyde-3-phosphate dehydrogenase (GAPDH) using the 2^ -ΔΔCT method. The qPCR primers, which were designed with PrimerBank [61], are listed in Table S2.

Statistical analysis

Data represent mean ± standard error of the mean. For parametric variables, the unpaired two-tailed Student t-test was used to assess the differences in mean values between two groups. For three groups, statistical analysis was performed with ANOVA with Tukey post hoc test. For nonparametric variables, the statistical significance of the differences was evaluated by the Mann–Whitney test or Kruskal–Wallis test. For the IPGTT, two-way ANOVA was performed for the evolution of blood glucose levels with a post hoc test using Bonferroni method. P < 0.05 was considered statistically significant.

Authors’ contributions

BLZ and YFH conceived and designed the experiments. XFS, NL, and BXL performed the experiments. LZ, FL, and YNH performed the bioinformatics analysis. XFS, YQF, and YM performed the statistical analysis and interpreted the data. XFS, LZ, FL, YX, and JL wrote the initial manuscript. BLZ and YFH reviewed and edited the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors have declared no competing interests.

Acknowledgements

This work was supported by the National Basic Research Program of China (Grant No. 2015CB554200), the Key Research Program of the Chinese Academy of Sciences (Grant No. KFZD-SW-219), and the National Natural Science Foundation of China (Grant Nos. 31601081 and 31471203).

Supplementary material

Supplementary data to this article can be found online at https://doi.org/10.1016/j.gphb.2019.03.001.

References


Carvalho BM, Saad MJ. Influence of gut microbiota on subclinical inflammation and insulin resistance. Mediators Inflamm 2013;2013:986734.


